

Experiment 21

Solubility

Pre-Lab Assignment

Before coming to lab:

- Read the lab thoroughly.
- Answer the pre-lab questions that appear at the end of this lab exercise.

Purpose

A series of standardized dilutions of copper(II) sulfate pentahydrate ($\text{CuSO}_4 \cdot 5 \text{H}_2\text{O}$) will be used to prepare a Beer's Law plot with spectrometry to find the relationship between concentration and absorbance. The average volume of 10 drops dispensed by a plastic pipette will be calibrated. With this, a series of solutions of $\text{CuSO}_4 \cdot 5 \text{H}_2\text{O}(\text{aq})$ will be prepared at different temperatures to determine whether the relationship between solubility and temperature is inverse or direct.

Background

Solubility is the measure of how much of a substance will dissolve in a set amount of solvent. Compounds that remain as solids in aqueous solution are classified as insoluble, whereas compounds that dissolve into separate ions in aqueous solution are classified as soluble. The degree to which a compound dissolves in aqueous solution is known as its solubility and is dependent on a number of factors, including temperature, concentration, and pH.

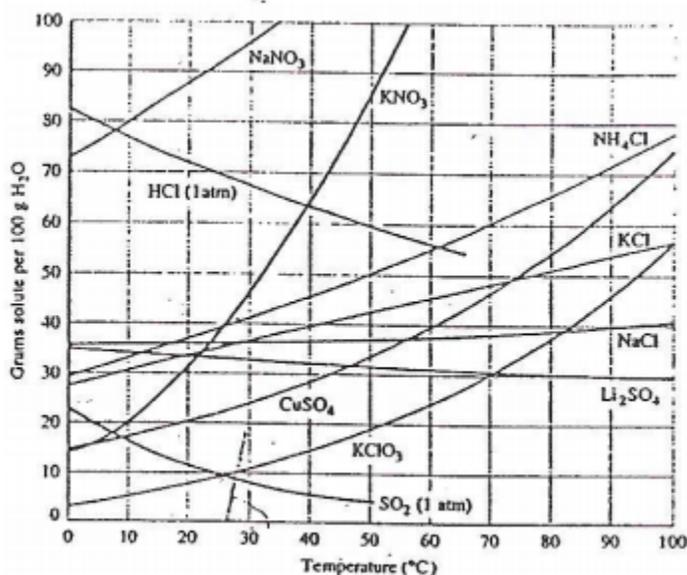


Fig. 1: Solubility of solids and gases at different temperatures

In general, the solubility of solids increases with increasing temperature whereas the solubility of gases decreases with increasing temperature (Fig. 1).

The solubility of a compound can be measured in terms of its concentration, and its concentration can be measured via spectrometry. In a spectrometer, a beam of light is shined onto the solution which absorbs some of its intensity while the rest of the light passes through. The instrument measures the ratio of light allowed to pass through the sample against its original intensity, giving readings in units of percent transmittance (%T). Since percent transmittance is based on an arbitrary scale of 0-100%, it is usually converted into absorbance (A) by Eqn. 2. Absorbance is a decimal and a unitless value.

$$A = -\log(\%T/100) \quad \text{Eqn. 2}$$

Beer's Law states that the absorbance of a solution is related to its concentration by Eqn. 3.

$$A = \epsilon l C \quad \text{Eqn. 3}$$

Here, ϵ is the molar absorptivity constant (in $M^{-1} \text{ cm}^{-1}$) which is individual to the identity of the solution, l is the path length of the light source inside the spectrometer (usually the width of the sample container, in cm) and C is the concentration of the solution (in M). Since ϵ and l are constant so long as the same compound and same spectrometer are used, a plot of concentration versus absorbance should be linear, giving a mathematical equation that can be used to determine the concentration of an unknown solution from its absorbance.

Example Problem: Determining the Concentration of an Unknown Solution

A previous set of standardized solutions of $\text{CuSO}_4 \cdot 5 \text{ H}_2\text{O}(\text{aq})$ gave a trendline for absorbance (y-axis) versus concentration (x-axis) of $y = 0.8895x + 0.0717$. A new solution was made by adding 10 drops of a saturated $\text{CuSO}_4 \cdot 5 \text{ H}_2\text{O}(\text{aq})$ from a calibrated plastic pipette to 2.50 mL of deionized water. Its absorbance was measured to be 0.354. The pipette was calibrated to dispense an average of 0.534 mL per 10 drops. Calculate the concentration of the original 10 drops of saturated solution.

Step 1: Use the trendline equation to convert absorbance to the concentration of the new solution

$$0.354 = 0.8895x + 0.0717$$

$$0.426 = 0.8895x$$

$$x = 0.479 \text{ M}$$

Step 2: Use $M_1V_1 = M_2V_2$ to find the original concentration

$$(0.479 \text{ M})(2.50 \text{ mL} + 0.534 \text{ mL}) = (M_2)(0.534 \text{ mL})$$

$$M_2 = 2.72 \text{ M}$$

Procedure

Part I: Calibration of the Plastic Pipette

1. Weigh the smallest size clean, dry Erlenmeyer flask with stopper. Record the mass in your data sheet.
2. Fill a small beaker approximately half-full with deionized water.
3. Fill your plastic pipette approximately a quarter full with the deionized water from the beaker in Step 2. Hold the dropper vertically and practice dispensing and counting single drops of liquid from it.
4. Once confident, dispense exactly 10 drops of deionized water into the empty Erlenmeyer flask from Step 1 and stopper it. Record the mass in your data sheet.
5. Add exactly 10 more drops of deionized water to the Erlenmeyer flask and stopper it. Record the new mass in your data sheet.
6. Add exactly 10 more drops of deionized water to the Erlenmeyer flask and stopper it. Record this third mass in your data sheet.
7. Record the temperature of the sample of deionized water used with a glass thermometer.
8. Squirt the pipette dry for later use.
9. Look up the accurate density of water at the temperature you recorded in Step 7.
10. Find the masses of each individual 10-drop portion of deionized water. Use the density you researched in Step 9 to convert each 10-drop mass to volume. Find the average volume of 10 drops dispensed from your plastic pipette.

Part II: Determination of Standard Solutions

1. Obtain six clean, dry small test tubes with rubber stoppers. Use a pencil to number the test tubes 1-6 on the white marking spots.
2. Use a pencil to label a 250-mL beaker "Waste" on the white (or blue) marking spot.
3. Clean and dry two small beakers (150-, 100-, or 50-mL capacity; smaller is better). Label one beaker "DI Water" and fill it with approximately 30 mL of deionized water. Label the other small beaker "CuSO₄ stock" and fill it with approximately 30 mL of 0.200 M CuSO₄·5 H₂O(aq).
4. Obtain a pipette bulb and set a volumetric pipets. You should have one volumetric pipette in each of the following volumes: 1 mL, 2 mL, 3 mL, and 5 mL. (There are no 4-mL pipettes.)
5. Fill each of the test tubes with the following:

#1: approximately 3 mL of deionized water. This will serve as your "blank" later.

#2: approximately 3 mL of 0.200 M CuSO₄·5 H₂O(aq)

Note: Do not use a graduated cylinder for the following measurements. Use a volumetric pipette. The pipette has higher precision, which is needed for this part of the experiment.

Note: Before using a volumetric pipette to measure out a liquid, rinse the pipette three times with the liquid you want to measure. Drain the rinses into the waste beaker.

#3: 4.00 mL of 0.200 M CuSO₄·5 H₂O(aq) + 1.00 mL deionized water

#4: 3.00 mL of 0.200 M CuSO₄·5 H₂O(aq) + 2.00 mL deionized water

#5: 2.00 mL of 0.200 M CuSO₄·5 H₂O(aq) + 3.00 mL deionized water

#6: 1.00 mL of 0.200 M CuSO₄·5 H₂O(aq) + 4.00 mL deionized water

6. Seal test tubes #3 through #6 with rubber stoppers and mix each one well by inverting the test tube a few times.
7. Calculate the concentration in Molarity of the solutions in Tubes #3-6.
8. Obtain a SpectroVis Spectrophotometer kit. The kit will include a spectrometer, a USB cable, and two small plastic cuvettes with caps. Do not lose these.

Log in to a lab computer and launch LoggerPro (there is a shortcut on the desktop). After Logger Pro has loaded, use the USB cable to connect the spectrometer to the mini-PC mounted on the back of the monitor. Logger Pro should automatically detect the spectrometer and enter data collection mode.

- To calibrate the spectrophotometer, go to the Experiment menu and select Calibrate, then Spectrometer (Fig. 2).

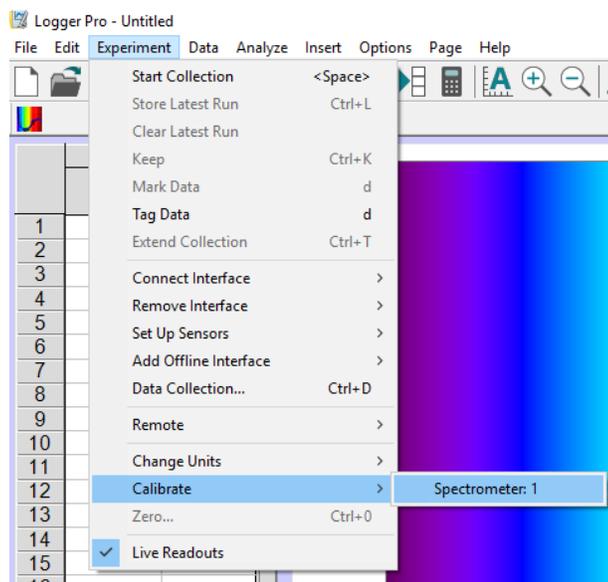


Fig. 2: Spectrometer Calibration

- Fill a plastic cuvette about three-fourths full of the solution from Tube #1 (your blank). Cap it. Remove any bubbles by tapping the cuvette on the workbench. Clean the smooth sides of the cuvette with a Kimwipe to remove fingerprints and dust, and then insert it into the sample holder in the spectrometer. Make sure that the smooth side is facing the light's path, indicated by the triangles on opposite sides of the sample holder.
- Follow the instructions in the dialog box to complete the calibration and then press "OK."
- Empty and dry the cuvette. Fill it about three-fourths full of the solution from Tube #2. Cap it. Remove any bubbles by tapping the cuvette on the workbench. Clean the smooth sides of the cuvette with a Kimwipe and then insert it into the sample holder in the spectrometer.
- Click "Collect." A full spectrum will be recorded across 400-750 nm. Click "Stop" to end data collection.
- Click the "Configure Spectrometer Data Collection" button (Fig. 3).

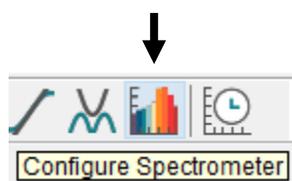


Fig. 1: Configure Spectrometer button

15. In the new window (Fig. 4), select "Absorbance vs. Concentration" as the collection mode. The wavelength of maximum absorbance (λ_{\max}) will be automatically selected from your previous graph from the curve's maximum. This number should be close to 680 nm. If it is not, click "Clear Selection" and type in 680 or select the wavelength closest to 680 nm. Otherwise, click "OK." You do not need to store the latest run.

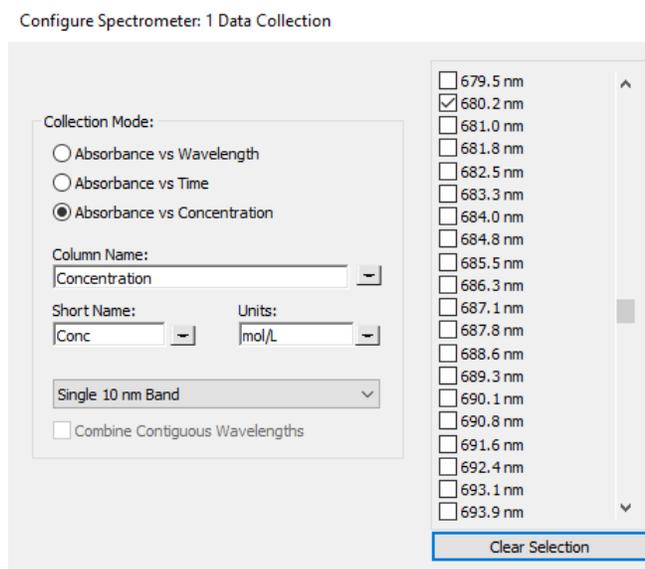


Fig. 2: Configure Spectrometer menu, select Collection Mode "Absorbance vs Concentration" and $\lambda_{\max}=680$ nm

16. With the solution from Tube #2 still in the sample holder, press the "Collect" button then press the "Keep" button. When prompted, enter the concentration of the sample – this is 0.200 M for Tube #2. Click "OK." Record the absorbance in Part II table on your data sheet. The absorbance is displayed in the bottom left corner of the screen.
17. Discard the solution in the cuvette in the waste beaker. Rinse the cuvette with deionized water and then thoroughly dry it.
18. Fill the dry cuvette about three-fourths full of the solution from Tube #3. Cap it. Remove any bubbles by tapping the cuvette on the workbench. Clean the smooth sides of the cuvette with a Kimwipe and then insert it into the sample holder in the spectrometer.
19. After the absorbance reading stabilizes, press "Keep." When prompted, enter the concentration of the sample, and click "OK." Record the absorbance on your data sheet.
20. Discard the solution in the cuvette in the waste beaker. Rinse the cuvette with deionized water and then thoroughly dry it.
21. Repeat Steps 18 – 20 with the remaining known solutions in Tubes #4-6.
22. Click "Stop" to end data collection.

23. Click the "Linear Fit" button. The equation for the trendline will be automatically displayed on the graph. Record the trendline equation on your data sheet.
24. Print the graph and include it with your data sheet.

Part III: Determination of Saturated Solutions

1. Obtain seven clean, dry small test tubes.
2. Add approximately 0.8-0.9 g of $\text{CuSO}_4 \cdot 5 \text{H}_2\text{O}(\text{s})$ crystals and 1.5 mL of deionized water to Tube #1.
3. Using a volumetric pipette, measure 2.50 mL of deionized water into Tubes #2-6.
4. Add approximately 3 mL of deionized water to Tube #7. This will be your "blank".
5. Prepare an ice bath by filling a large beaker approximately 2/3 full of ice and tap water.
6. Place Tube #1 in the ice bath in Step 5. Stir the ice-water mixture with a glass stirring rod while monitoring the temperature with a glass thermometer. When the temperature ceases to decrease, record it in your data sheet. Wipe the stirring rod dry before using it to stir the mixture inside Tube #1 for three minutes. Keep the tube in the ice bath and let it settle for at least one minute.
7. Using the same plastic pipette from Part I, carefully transfer exactly 10 drops from the top of the saturated solution in Tube #1 to Tube #2. Squirt any excess solution left in the pipette back into Tube #1.
8. Discard the ice bath in the sink and fill the beaker approximately two-thirds full of room temperature tap water. Put Tube #1 back into the water bath. Stir the water bath with a glass stirring rod for at least three minutes. Wipe the stirring rod dry before using it to stir the mixture inside Tube #1 for about three minutes. Record the temperature of the water bath on your data sheet.
9. Allow the mixture inside Tube #1 to settle for at least one minute. Using the same plastic pipette from Part I, transfer exactly 10 drops from the top of the saturated solution Tube #1 to Tube #3. Squirt any excess solution left in the pipette back into Tube #1.
10. Gather a Bunsen burner, iron ring, and wire gauze. Set the beaker with the water bath and Tube #1 carefully on top of the burner and iron ring.
11. Heat the water bath and Tube #1 to approximately 40°C. Stir the water bath with a glass stirring rod for at least three minutes. Wipe the stirring rod dry before using it to stir the mixture inside Tube #1 for about three minutes. Record the temperature of the water bath on your data sheet.
12. Allow the mixture inside Tube #1 to settle for at least one minute. Using the same plastic pipette from Part I, transfer exactly 10 drops from the top of the Tube #1 to Tube #4. Squirt any excess solution left in the pipette back into Tube #1.

13. Repeat Steps 11-12 twice more, heating the water bath to approximately 60°C and then 80°C.
If you run out of liquid in Tube #1, add approximately 0.50 mL of deionized water.
If you run out of crystals in Tube #1, add approximately 0.5 g $\text{CuSO}_4 \cdot 5 \text{H}_2\text{O}(\text{s})$.
14. Shake Tubes #2-6 gently.
15. Repeat Part I Steps 4-17 to set up the spectrophotometer. Use Tube #7 as your "blank" to calibrate the spectrometer. Use stock solution (0.200 M $\text{CuSO}_4 \cdot 5 \text{H}_2\text{O}$) to find λ_{max} , which should still be around 680 nm.
16. Change the collection mode via the Configure Spectrometer button to Absorbance vs. Concentration. Record the absorbance for Tubes #2-6 on your data sheet. When asked for concentration, enter the tube number. Do not print this graph.
17. Dispose of Tubes #1-6 in the appropriate waste container.
18. Using the trendline equation from the graph in Part II, convert each absorbance to concentration, in M, for Tubes #2-6 for the dilute solutions.
25. Using $M_1V_1 = M_2V_2$, convert the dilute solution concentrations in Step 18 to the saturated solution concentrations from the original 10 drops removed from Tube #1.
26. Plot a graph of concentration of saturated solutions (y-axis) vs temperature (x-axis). Determine whether the relationship between solubility and temperature is direct or inverse. Print out the graph and include it with your post-lab assignment.

Experiment 21—Data Sheet

Name: _____

Part I: Calibration of the Plastic Pipette

| Number of Drops | Total Mass (g) | Mass of 10 Drops (g) | Volume of 10 Drops (mL) |
|-----------------|----------------|----------------------|-------------------------|
| 0 | | | |
| 10 | | | |
| 20 | | | |
| 30 | | | |

1. Temperature of deionized water ($^{\circ}\text{C}$) _____

2. Density of water at this temperature from CRC (g/mL) _____
show calculations for one 10 drop portion:

3. Average volume of 10 drops (mL) _____
show calculation:

Part II: Determination of Standard Solutions

| Tube # | mL of 0.200 M CuSO₄·5 H₂O added | mL H₂O added | M CuSO₄ diluted | Absorbance |
|---------------|--|--------------------------------|---------------------------------------|-------------------|
| 2 | about 3 | 0 | 0.200 | |
| 3 | 4.00 | 1.00 | | |
| 4 | 3.00 | 2.00 | | |
| 5 | 2.00 | 3.00 | | |
| 6 | 1.00 | 4.00 | | |

show calculation for Tube #3 diluted concentration:

1. Trendline Equation from graph:

Part III: Determination of Saturated Solutions

| Tube # | Temperature (°C) | Absorbance | Molarity (dilute) | Molarity (saturated) |
|---------------|-------------------------|-------------------|--------------------------|-----------------------------|
| 2 | | | | |
| 3 | | | | |
| 4 | | | | |
| 5 | | | | |
| 6 | | | | |

Show calculation for Tube #2, Molarity (dilute) (use graph equation):

Show calculation for Tube #2, Molarity (saturated):

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5. The CRC Handbook value for the solubility of $\text{CuSO}_4 \cdot 5 \text{H}_2\text{O}$ at 25°C is 22.0 g per 100 g water. Calculate the percent error for experiment value for the solubility from the previous question.

6. Give three experimental reasons why your solubility value may be different than the CRC's.

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